

1 ***In vitro* sensitivity of *Sclerotinia sclerotiorum* isolated from soybeans to**  
2 **fungicides**

3 Ricardo Brustolin<sup>1,3</sup> e Erlei Melo Reis<sup>2</sup>

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5 <sup>1</sup>Universidade de Passo Fundo, <sup>3</sup>endereço atual Pioneer sementes, Passo Fundo, RS

6 <sup>2</sup> OR Melhoria de Sementes Ltda, Passo Fundo, RS.

7 Autor para correspondência: Erlei Melo Reis ([erleireis@upf.br](mailto:erleireis@upf.br)).

8 Data de chegada: 00/00/2015. Aceito para publicação em: 00/00/2015

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10 **ABSTRACT**

11 Brustolin, R.; Reis, E. M. *In vitro* sensitivity of *Sclerotinia sclerotiorum* isolated from  
12 soybeans to fungicides. **Summa Phytopathologica**, v. n. p. 00-00, 2015.

13 *In vitro* experiments conducted in the laboratory, the sensitivity of *Sclerotinia*  
14 *sclerotiorum* isolated from soybean, to boscalid, carbendazim, fluazinam, procymidone,  
15 tebuconazole and thiophanate methyl was determined. To found the 50% inhibitory  
16 concentration (IC<sub>50</sub>) of mycelial growth, the potato-sucrose agar medium was  
17 supplemented with 0.0, 0.01, 0.10, 1.0, 25.0 and 50.0 mg/L of active ingredient, poured  
18 into petri dishes and observing four replications. Discs containing mycelium and  
19 substrate, 6.0 mm in diameter, were deposited in the dishes center and kept for seven days  
20 in a growth chamber at 18 ± 2°C and 12 h photoperiod. The chemical effect on the  
21 mycelial growth was determined by measuring the colonies diameter. The data, as percent  
22 inhibition relative to the concentration zero, were submitted to logarithmical regression  
23 and with the generated equation, the IC<sub>50</sub> was calculated. IC<sub>50</sub> ranged from <0.01 for the  
24 fluazinam, 0.09 for carbendazim, 0.13 for tebuconazole, 0.26 for procymidone, 0.29 for  
25 boscalida and 42 mg/L for thiophanate methyl. It was confirmed that fluazinam, a  
26 multisite, protectant was the most potent fungicide against *S. sclerotiorum*.

27 **Keywords** – Fluazinam, *Glycine max*, white mold, mycelial sensitivity.

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29 **RESUMO**

1 Brustolin, R.; Reis, E. M. Sensibilidade *in vitro* de *Sclerotinia sclerotiorum*, isolados de  
2 soja, a fungicidas. **Summa Phytopathologica**, v. n. p. 00-00, 2015.

3 Em experimentos conduzidos *in vitro* em laboratório, determinou-se a  
4 sensibilidade de *Sclerotinia sclerotiorum*, isolados de soja, aos fungicidas boscalida,  
5 carbendazim, fluazinam, procimidona, tebuconazol e tiofanato metílico. Para a  
6 determinação da concentração inibitória de 50% (CI<sub>50</sub>) do crescimento micelial, o meio  
7 de cultura de batata-sacarose-ágar foi suplementado com 0,0; 0,01; 0,10; 1,0; 25,0 e 50,0  
8 mg/L de ingrediente ativo, vertido em placas de Petry e observando-se quatro repetições.  
9 Discos contendo micélio e substrato, de 6,0 mm de diâmetro, foram depositados no centro  
10 dos recipientes e mantidos por sete dias em câmara de crescimento a 18 ± 2°C e  
11 fotoperíodo de 12h. O efeito dos tratamentos sobre o crescimento micelial foi  
12 determinado pela mensuração do diâmetro das colônias. Os dados da percentagem de  
13 inibição em relação à concentração zero, foram submetidos a regressão logarítmica e com  
14 a equação gerada calculada a CI<sub>50</sub>. A CI<sub>50</sub> variou de < 0,01 para o fluazinam, 0,09 para o  
15 Carbendazim, 0,13 para o tebuconazol, 0,26 para a procimidona, 0,29 para a boscalida e  
16 0,42 mg/L para o tiofanato metílico. Confirmou-se que o fungicida multissítio, protetor  
17 mais potente inibidor do micélio de *S. sclerotiorum* foi o fluazinam.

18 **Palavras-chave** – Fluazinam, *Glycine max*, mofo branco, sensibilidade micelial.

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## INTRODUCTION

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22 The Brazilian soybean crop [*Glycine max* (L.) Merr] cultivated in 2013-14  
23 covered an area of 29.8 million hectares and yielded 2.8 t/ha, resulting in an overall  
24 production of 90 million tons (3).

25 *Sclerotinia* stem rot (SSR), or white mold (WM) of soybean, is a significant yield-  
26 limiting disease in Brazil. This disease, caused by the fungus *Sclerotinia sclerotiorum*  
27 (Lib.) de Bary is among the most nonspecific, omnivorous, and successful of plant  
28 pathogens. Plants susceptible to this pathogen reach 652 species (nt.ars-  
29 grin.gov/fungalatabases/index.cfm).

30 WM varies in incidence and severity from season to season according to weather  
31 conditions. Considering white mold area with the highest incidence, the estimated

1 infested acreage in 2008 was 4%, rising to 6.5% in 2009, 9.8% in 2010, 14.4% in 2011,  
2 22.9% in 2012 and reaching 25.5% in 2013 (11).

3 Soybean cultivars with resistance/tolerance to Ss are not yet available and the  
4 soybean area is grown under monoculture which contributes to increase the sclerotia bank  
5 in the soil, season after season. Crop rotation with nonsusceptible crops could reduce the  
6 inoculum in the soil of cultivated area but it is not followed by growers (2).

7 Losses caused by SSR can be appraised by the damage function  $y = - 6.7 x +$   
8 1,000, where y represents grain yield normalized to 1,000 kg/ha and x represents SSR  
9 incidence in plants (13).

10 An effective disease management strategy integrates several tactics that include  
11 cultural practices, as chemical and biological control. However, chemical is the most  
12 used.

13 Boscalid, carbendazim, iprodione, fluazinam, fludioxonil, procymidone,  
14 tiophanate methyl, and vinclozolin, are among the most used fungicides to control SSR  
15 especially in canola, edible beans, peanuts and soybean (7, 8, 9, 10, 12, 18).

16 The objective of this work was to identify the most potent fungicides among those  
17 reported with the highest fungitoxicity towards *S. sclerotiorum* isolates to be used in an  
18 antiresistance strategy.

19

20

## MATERIAL AND METHODS

21 **Sclerotinia isolates.** To evaluate the mycelial growth of Ss to fungicides, the  
22 chemicals were incorporated in an agar medium, similar to the method described by  
23 Russel (5). Five selected monoascosporic isolates (CX, from Coxilha county Rio Grande  
24 do Sul state, SC, from Santa Catarina state, BA, from Luiz Eduardo Magalhães county  
25 Bahia state, PR, from Faxinal county Paraná state, and GO, from Rio Verde county,  
26 Goiás state) were preserved in test tubes with PSA (40 g potatoes, 10 g sucrose, 14 g agar  
27 in 1,000 mL distilled water) medium in a refrigerator at 5°C and used throughout this  
28 work.

29 **Fungicides.** Six chemicals, containing carbendazim (Bendazol 500 SC),  
30 fluazinam (Frowncide 500 SC), boscalid (Cantus 500 WG), procymidone (Sumilex 500  
31 WP), and tebuconazole (Folicur 200 EC), were assessed for Ss mycelial growth  
32 inhibition.



1 Table 1. Isolate, equation, coefficient of determination ( $r^2$ ), fungicide concentration to  
 2 inhibit 50% ( $IC_{50}$ ) the mycelial growth of *Sclerotinia sclerotiorum* by  
 3 fluazinam

Isolate	Equation*	$r^2$	$IC_{50}^{**}$	$p$
CX	$y=2.27\ln(x)+93.5$	0.84	<0.01	<0.0001
SC	$y=2.79\ln(x)+92.45$	0.75	<0.01	<0.0001
BA	$y=3.44\ln(x)+90.96$	0.73	<0.01	<0.0001
PR	$y=2.53\ln(x)+92.58$	0.80	<0.01	<0.0001
GO	$y=3.40\ln(x)+91.14$	0.73	<0.01	<0.0001
Media	$y=2.89\ln(x)+92.13$	0.76	<0.01	<0.0001

4 \*y = percent mycelial growth inhibition. \*\*Concentration (mg/L) calculated by the equation.

5

6 For carbendazim, a benzimidazole, site specific, penetrant, mobile fungicide,  $IC_{50}$   
 7 ranged from 0.05, PR isolate to 0.14 mg/L, BA isolate and  $r^2$  ranged from 0.67 to 0.80  
 8 with a mean of 0.09 mg/L (Table 2). Wang et al (24) and Yong et al. (22) have reported  
 9 the Ss resistance to carbendazim a high risk fungicide belonging to the benzimidazole  
 10 group such as tiophanate methyl and benomyl (6).

11 Although showing a high fungitoxicity towards Ss, under field application should  
 12 be take into consideration the sensitivity loss by *Corynespora cassiicola* (Berk. & M.A.  
 13 Curtis) Wei, isolates from soybean, to carbendazim (1).

14

15 Table 2. Isolates, equation, coefficient of determination ( $r^2$ ), fungicide concentration to  
 16 inhibit 50% ( $IC_{50}$ ) the mycelial growth of *Sclerotinia sclerotiorum* by  
 17 carbendazim

Isolate	Equation*	$r^2$	$CI_{50}^{**}$	$p$
CX	$y=10.01\ln(x)+70.87$	0.76	0.12	<0.0001
SC	$y=9.02\ln(x)+72.61$	0.67	0.08	<0.0001
BA	$y=9.78\ln(x)+69.48$	0.75	0.14	<0.0001
PR	$y=7.81\ln(x)+73.38$	0.80	0.05	<0.0001
GO	$y=9.22\ln(x)+71.66$	0.67	0.10	<0.0001
Media	$y=9.17\ln(x)+71.6$	0.73	0.09	<0.0001

18 \*y = percent mycelial growth inhibition. \*\*Concentration (mg/L) calculated by the equation.

19

20 For the DMI site specific, penetrant, mobile, tebuconazole,  $IC_{50}$  ranged from  
 21 <0.01, for CX isolate to 0.87 mg/L, SC isolate and with a mean of 0.13. In addition,  $r^2$   
 22 ranged from 0.67 to 0.95 (Table 3). This chemical should not use alone in soybean due to  
 23 its sensitivity reduction towards *Phakopsora pachyrhizi* Sydow (14, 17).

1 Table 3. Isolates, equation, coefficient of determination ( $r^2$ ), fungicide concentration to  
 2 inhibit 50% ( $IC_{50}$ ) the mycelial growth of *Sclerotinia sclerotiorum* by  
 3 tebuconazole

Isolado	Equação*	$r^2$	$CI_{50}^{**}$	$p$
CX	$y=5.09\ln(x)+87.25$	0.67	<0.01	<0.0001
SC	$y=13.10\ln(x)+51.81$	0.92	0.87	<0.0001
BA	$y=11.58\ln(x)+63.73$	0.95	0.31	<0.0001
PR	$y=8.50\ln(x)+76.21$	0.82	0.05	<0.0001
GO	$y=9.77\ln(x)+68.21$	0.91	0.16	<0.0001
Media	$y=9.61\ln(x)+69.44$	0.94	0.13	<0.0001

4 \*y = percent mycelial growth inhibition. \*\*Concentration (mg/L) calculated by the equation.

5

6 Regarding procymidone, a dicarboxamide, nonpenetrant, protectant fungicide,  
 7  $IC_{50}$  ranged from 0.15, SC isolate to 0.40 mg/L for BA isolate, with a mean of 0.26 mg/L,  
 8 and  $r^2$  ranged from 0.81 to 0.88 (Table 4). The activity of dicarboximide fungicides was  
 9 first reported in the early 1970's with the three key commercial products being introduced  
 10 within three years; iprodione, vinclozolin and procymidone.

11

12 Table 4. Isolates, equation, coefficient of determination ( $r^2$ ), fungicide concentration to  
 13 inhibit 50% ( $IC_{50}$ ) the mycelial growth of *Sclerotinia sclerotiorum* by  
 14 procymidone

Isolado	Equação*	$r^2$	$CI_{50}^{**}$	$p$
CX	$y=10.15\ln(x)+71.38$	0.81	0.20	<0.0001
SC	$y=10.35\ln(x)+69.87$	0.85	0.15	<0.0001
BA	$y=13.15\ln(x)+62.03$	0.85	0.40	<0.0001
PR	$y=12.46\ln(x)+62.61$	0.88	0.36	<0.0001
GO	$y=12.83\ln(x)+63.06$	0.85	0.36	<0.0001
Media	$y=11.79\ln(x)+65.70$	0.87	0.26	<0.0001

15 \*y = percent mycelial growth inhibition. \*\*Concentration (mg/L) calculated by the equation

16

17 Considering boscalid, a site specific carboxamide fungicide, nonpenetrant,  
 18 protectant fungicide,  $IC_{50}$  ranged from 0.11, PR isolate to 0.57 mg/L, GO isolate, with a  
 19 mean of 0.29, and  $r^2$  ranged from 0.73 to 0.84 (Table 5). The  $IC_{50}$  found by Xin Liu et al.  
 20 (21) for boscalid ranged from 0.002 to 0.391 mg/L with a mean of 0.042 mg/L, 6.0-fold  
 21 lower than we found.

1 Table 5. Isolates, equation, coefficient of determination ( $r^2$ ), fungicide concentration to  
 2 inhibit 50% ( $IC_{50}$ ) the mycelial growth of *Sclerotinia sclerotiorum* by boscalid

Isolate	Equation*	$r^2$	$IC_{50}^{**}$	$p$
CX	$y=10.31\ln(x)+60.38$	0.94	0.37	<0.0001
SC	$y=9.26\ln(x)+60.66$	0.88	0.32	<00001
BA	$y=8.86\ln(x)+63.07$	0.93	0.23	<0.0001
PR	$y=8.20\ln(x)+68.45$	0.89	0.11	<0.0001
GO	$y=9.04\ln(x)+55.04$	0.93	0.57	<0.0001
Mean	$y=9.22\ln(x)+61.52$	0.96	0.29	<0.0001

3 \*y = percent mycelial growth inhibition. \*\*Concentration (mg/L) calculated by the equation.

4  
 5 In relation to tiophanate methyl, also a benzimidazole,  $IC_{50}$  ranged from 0.19, BA  
 6 isolate to 0.54 mg/L, GO isolate, with a mean of 0.42, and  $r^2$  ranged from 0.83 to 0.90  
 7 (Table 6). The first case of Ss resistant to benomyl, also a benzimidazol, was reported by  
 8 Gossen et. al. (6).

9  
 10 Table 6. Isolates, equation, coefficient of determination ( $r^2$ ), fungicide concentration to  
 11 inhibit 50% ( $IC_{50}$ ) the mycelial growth of *Sclerotinia sclerotiorum* by tiophanate  
 12 methyl

Isolado	Equação*	$r^2$	$IC_{50}^{**}$	$p$
CX	$y=12.40\ln(x)+59.63$	0.90	0.46	<0.0001
SC	$y=12.21\ln(x)+58.07$	0.86	0.52	<0.0001
BA	$y=10.31\ln(x)+67.32$	0.85	0.19	0.0011
PR	$y=10.97\ln(x)+57.64$	0.86	0.50	<0.0001
GO	$y=12.74\ln(x)+57.82$	0.83	0.54	<0.0001
Media	$y=11.73\ln(x)+60.1$	0.86	0.42	<0.0001

13 \*y = percent mycelial growth inhibition. \*\*Concentration (mg/L) calculated by the equation

14  
 15 Xin et al. (21) determined for iprodione  $IC_{50}$  ranged from 0.163 to 0.734  $\mu\text{g/ml}$   
 16 with a mean of 0.428  $\mu\text{g/ml}$  similar to our findings for tiophanate methyl.

17 The overall mean for fungitoxicity ranged from <0.01 for fluazinam, the most  
 18 fungitoxic, to 0.42 mg/L for tiophanate methyl the least potent, and  $r^2$  ranged from 0.73  
 19 to 0.96 (Table 7). Figueiredo reported tiophanate methyl as highly toxic but at 1 mg/L, a  
 20 concentration 2.38-fold higher than the concentration here tested.

1 Table 7. Fungicide, equation and fungicide concentration to inhibit 50% (IC<sub>50</sub>) mycelial  
 2 growth of *Sclerotinia sclerotiorum*, means of five isolates

Fungicide	Equation*	r <sup>2</sup>	CI <sub>50</sub> **	p**
Fluazinan	y=2.89ln(x)+92.13	0.76	< <b>0.01</b>	<0.0001
Carbendazim	y=9.17ln(x)+71.6	0.73	<b>0.09</b>	<0.0001
Tebuconazole	y=9.61ln(x)+69.44	0.94	<b>0.13</b>	<0.0001
Procymidone	y=11.79ln(x)+65.70	0.87	<b>0.26</b>	<0,0001
Boscalid	y=9.22ln(x)+61.52	0.96	<b>0.29</b>	<0.0001
Tiophanate methyl	y=11.73ln(x)+60.1	0.86	<b>0.42</b>	<0.0001

3 \*y = percent mycelial growth inhibition. \*\*Concentration (mg/L) calculated by the equation.  
 4  
 5

6 According to Edgington et al (4) all isolates were highly sensitive to the tested  
 7 fungicides with CI<sub>50</sub> < 1.0 mg/L.

8 Although not tested in our work, Mueller et al. (12) reported vinclozolin as the  
 9 fungicide showing the highest fungitoxicity towards Ss. Kuang et al. (9), found the mean  
 10 IC<sub>50</sub> values for fludioxonil based on inhibition of mycelial growth of 120 wild-type  
 11 isolates, 0.015 ± 0.005 µg/ml. Positive cross-resistance was not detected between  
 12 fludioxonil and benzimidazole fungicides but was detected between fludioxonil and  
 13 dicarboximide fungicides which are considered as high resistance risk fungicides by  
 14 FRAC (5).

15 The search for powerful fungicide to Ss control should be a continuous task.  
 16 Various fungicides have shown high fungitoxicity to Ss, however, under field conditions,  
 17 exhibit reduced efficiency. This difference can be attributed to difficulty to protect the Ss  
 18 infection sites, the senesced petals in the lower third of the plant and protected by  
 19 innumerable layers of leaves.

20 Soybean cultivars with resistance/tolerance to Ss are not yet available. The WM  
 21 infested area continuous to grow, reaching 25.5% in 2013 growing season (11). This large  
 22 soybean area, grown under monoculture, which contributes to increase the sclerotia bank  
 23 in the soil, season after season. Crop rotation with nonsuseptible crops could help to  
 24 reduce the inoculum in the cultivated soil but it is not follow by growers (2).





- 1 *sclerotiorum*) em soja: safra 2008/2009. Curitiba: ABRATES, 2009. 3 p. (**Informativo**  
2 **Técnico, 19**).
- 3 8. Hisada, Y.; Maeda, K.; Tottori, N. & Kawase, Y. (1976). Plant disease control by N-  
4 (3,5-dichlophenyl)-1,1-dimethyl-cyclopropane-1,2-dicarboxamide. **Journal of**  
5 **Pesticide Science**, n. 1, p. 145-149, 1976.
- 6  
7  
8 9. Kuang, J.; Hou, Y-P.; Wang, J-X.; Zhou, M-X. Sensitivity of *Sclerotinia sclerotiorum*  
9 to fludioxonil: In vitro determination of baseline sensitivity and resistance risk. **Crop**  
10 **Protection, v. 30, n.7, p. 876-882, 2011.**
- 11 10. Matheron, M.A.; Porchas, M. Activity of boscalid, fenhexamid, fluazinam,  
12 fludioxonil, and vinclozolin on growth of *Sclerotinia minor* and *S. sclerotiorum* and  
13 development of lettuce drop. Plant disease, St. Paul, v. 88, n. 6  
14 p. 665 – 668, 2004.
- 15 11. Meyer, M. C.; Campos, H. D.; Godoy, C. V.; Utiamada, C. M. Ensaios cooperativos de  
16 controle químico de mofo branco na cultura da soja: safras 2009 a 2012. Londrina:  
17 Embrapa Soja, 2014. 100p. (Documentos / Embrapa Soja, ISSN 1516-781X; n.345).
- 18
- 19 12. Mueller, D. S.; Dorrance, A. E.; Derksen, R. C.; Ozkan, E.; Kurle, J. E.; Grau, C. R.;  
20 Gaska, J. M.; Hartman, G. L.; Bradley, C. A.; Pedersen, W. L. Efficacy of fungicides  
21 on *Sclerotinia sclerotiorum* and their potential for control of sclerotinia stem rot on  
22 soybean. **Plant Disease**, St Paul, v. 86, p. 26-31, 2002.
- 23 13. Reis, E. M.; Zanatta, M.; Hercules, H. D.; Silva, L. H, C. P.; Meyer, M. C.; Nunes 5  
24 Junior, J.; Pimenta, C. P.; Cassetari Neto, D.; Machado, A. Q.; Juliatti, F. C.; Utiamada,  
25 C. M. A critical-point model to appraise the damage caused to soybean yield by white  
26 mold. **Summa Phytopathologica**, Botucatu, 2014 (in press).
- 27
- 28 14. Reis, E. M.; Deuner, E. *In vivo* sensitivity reduction of *Phakopsora pachyrhizi* to  
29 tebuconazol. **Summa Phytopathologica**, Botucatu, 2015 (in press).
- 30
- 31 15. Russel, P. E. **Sensitivity baselines in fungicide resistance research and**  
32 **management**. Cambridge, FRAC Monograph no.3, 2004. 56p.
- 33

- 1 16. Shao, W.; Zhang, Y.; Ren, W.; Chen, C. Physiological and biochemical characteristics  
2 of laboratory induced mutants of *Botrytis cinerea* with resistance to fluazinam.  
3 **Pesticide Biochemistry and Physiology**, Cambridge, Available online 16 October  
4 2014.
- 5  
6 17. Silva, L. H. C. P.; Campos, H.D.; Silva, J.R.C. Eficácia reduzida de triazóis no controle  
7 da ferrugem asiática. **Fitopatologia Brasileira**, Brasília, D.F., v. 33, p. 228, 2008  
8 (Suplemento).
- 9  
10 18. Smith, F.D.; Phipps, P.M.; Stipes, R.J. Agar plate, soil plates, and field evaluation of  
11 fluazinam and other fungicides for control of *Sclerotinia minor* on peanuts. **Plant**  
12 **Disease**, St Paul, v. 75, p. 1138-1143, 1991.
- 13 18. Sumida, C.H.; Canteri, M.G.; Peitl, D.C.; Orsini., I.P.; Tibolla, F.; Araújo, FG.A.;  
14 Chagas, D.F. Inibição micelial *in vitro* de *Sclerotinia sclerotiorum* by fungicidas.  
15 **Summa Phytopathologica**, Botucatu, [online]. v.40, n.1, p. 90-91, 2014.
- 16 19. Vieira, R.R.; Pinto, Cleide M.F.; Paula Jr, Trazilbo J. Chemigation with benomyl and  
17 fluazinam and their fungicidal effects in soil for white mold control on dry beans.  
18 **Fitopatologia Brasileira**, Brasília, DF, v.28, n.3, p. 245-250, 2003.
- 19 20. XIN, L.; YANNI, Y.; LEIYAN, Y.; MICHAILIDES T.; ZHONGHUA, J.. Biochemistry and  
20 physiology sensitivity to iprodione and boscalid of *Sclerotinia sclerotiorum* isolates  
21 collected from rapeseed in China. **Pesticide biochemistry and**  
22 **physiology**, 2009, v. 95, n°2, p. 106-112, 2009.
- 23 21. Yong, W., Yi-Ping, H.; Chang-Jun. C.; Ming-Guo Z. Detection of resistance in  
24 *Sclerotinia sclerotiorum* to carbendazim and dimethachlor in Jiangsu Province of  
25 China. **Australasian Plant Pathology**, 2014, Volume 43, Issue 3, pp 307-312, 2014.  
26
- 27 22. Zancan. W. L.A.; Machado, J.C.; Sousa, B.F.M.; Matos, C.S.M. Crescimento micelial,  
28 produção e germinação de escleródios de *Sclerotinia sclerotiorum* na presença de  
29 fungicidas químicos e *Trichoderma harzianum*. **Biosciences Journal**, Uberlândia, v.  
30 28, n. 5, p.782-789, 2012.  
31

- 1 23. Wang, Y; Duan, Y.-B.; Zhou, M.-G. Molecular and biochemical characterization of  
2 boscalid resistance in laboratory mutants of *Sclerotinia sclerotiorum*. **Plant**  
3 **Pathology**, Article first published online: 18 JUN 2014 DOI: 10.1111/ppa.12246,  
4 2014.